

TRITERPENOID—IV¹

THE CYCLAMIGENINS A¹, A², C, AND D

R. Ó DORCHAÍ, H. E. RUBALCAVA and J. B. THOMSON

Department of Chemistry, University College, Dublin, Ireland

B. ZEEH

Chemisches Institut der Universität Tübingen, Germany

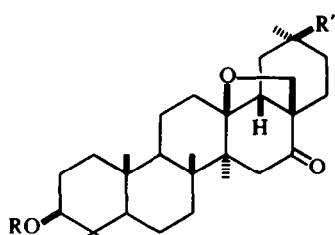
(Received in the UK 12 March 1968; accepted for publication 8 April 1968)

Abstract—The cyclamigenins A¹ and C are 30 β - (IXa) and 30 α - (IXb) ethoxy-28,30-epoxyolean-12-en-3 β ,16 α -diol, respectively. Cyclamigenin A² is 30(or 29)-noraegicerin (V) and cyclamigenin D is 30,30-dimethoxyaegicerin (VII). The identifications are based mainly on spectroscopic data but cyclamigenins A¹ and C have been converted to cyclamiretin D (XI) and cyclamigenin D has been converted to 3 β ,28-diacetoxy-16-keto-olean-12-en-30-oic acid (XV).

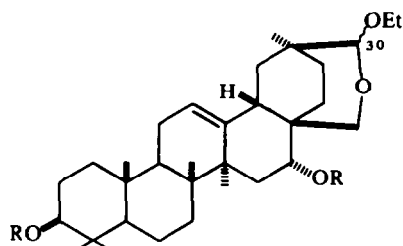
IN A previous communication¹ we described the isolation of the cyclamigenins A, B (I), C, D, and E from the amorphous saponin of *Cyclamen europaeum* corms. A sixth triterpene, which is intermediate in chromatographic mobility between cyclamigenins A and B, has now been isolated. The new cyclamigenin is designated A², the original A now being A¹. This paper describes the determination of the structures of cyclamigenins A¹, A², C, and D.

Cyclamigenins A¹ and C are isomers, the mass spectra of which show molecular ions at m/e 500 (C₃₂H₅₂O₄). They are stable to alkali and their IR spectra, which are almost identical, show no carbonyl bands but strong absorption at 3400 (O—H stretch) and 1000 cm⁻¹ (C—O stretch) and the group of peaks at ca. 820 cm⁻¹ characteristic² of the 12(13)-double bond in pentacyclic triterpenes. Both compounds show only end-absorption in the UV. Two OH groups are present in each compound, as is demonstrated by the ready formation of diacetates the mass spectra of which exhibit molecular ions at m/e 584 (C₃₆H₅₆O₆). That both of the remaining O atoms in cyclamigenins A¹ and C are present in ether functions is indicated by the absence of O—H stretching vibrations in the IR spectra of the acetates. A Zeisel estimation shows that one of these ether functions is an EtO group.

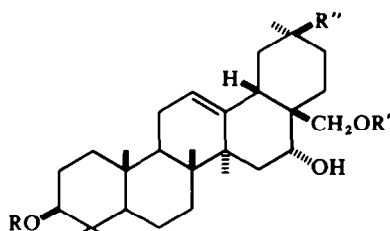
The most obvious structures for cyclamigenins A¹ and C are the 30-epimeric, mixed acetals (IX) of cyclamiretin D (XI).³ The NMR spectra of the acetates (X) are consistent with this hypothesis in that they show the presence of six angular Me groups; an EtO group as a triplet at ca. τ 8.75, partly obscured by the methylene envelope and an angular Me signal, and a quartet at ca. τ 6.45, partly obscured by the 28-methylene quartet centred at ca. τ 6.5 ($J = -12$ Hz); the H-16 β multiplet at ca. τ 6.4; the H-12 multiplet at τ 4.65; and H-30 as a singlet at τ 5.6. Confirmation of these structures was obtained by mild acid-hydrolysis of the acetates both of which yield cyclamiretin D monoacetate (XII), identical with a sample prepared by a similar hydrolysis of cyclamiretin D diacetate (XIII).^{3, 4}



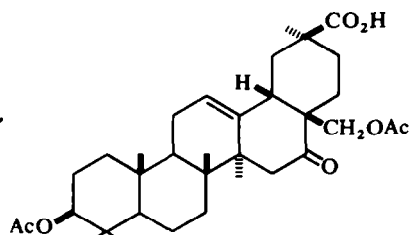
- I: R = H, R' = CHO
 II: R = Ac, R' = CHO
 III: R = H, R' = Me
 IV: R = Ac, R' = Me
 V: R = R' = H
 VI: R = Ac, R' = H
 VII: R = H, R' = CH(OMe)₂
 VIII: R = Ac, R' = CH(OMe)₂



- IXa: R = H, 30β
 IXb: R = H, 30α
 Xa: R = Ac, 30β
 Xb: R = Ac, 30α



- XI: R = R' = H, R'' = CHO
 XII: R = Ac, R' = H, R'' = CHO
 XIII: R = R' = Ac, R'' = CHO
 XIV: R = R' = H, R'' = CH(OEt)₂



XV

Ring E is necessarily a twist-boat in cyclamigenins A¹ and C, while the ease of acetylation of the 16α-OH group and the unusually high-field position of the H-16β resonance (τ 6.4) in the NMR spectra of the acetates (i.e. H-16β lies above the double bond) indicate that ring D is a boat with C-13 and C-16 bow and stern. Of the two possible conformations for the oxide bridge the *endo* form (Fig. 1) must be favoured as non-bonding interactions are much less severe than in the *exo* form (Fig. 2). Thus, in the *exo* form (Fig. 2) H-16β and H-28β, AcO-16α and H-28α, and Me-20 and EtO-30α (or H-30α) are eclipsed and in the axial epimer the oxygen of the EtO group is close to H-18 (1.5 Å from Dreiding models). In the *endo* form (Fig. 1) all of

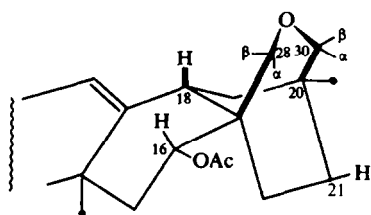


Fig. 1

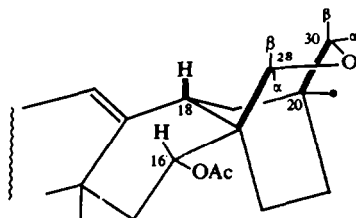
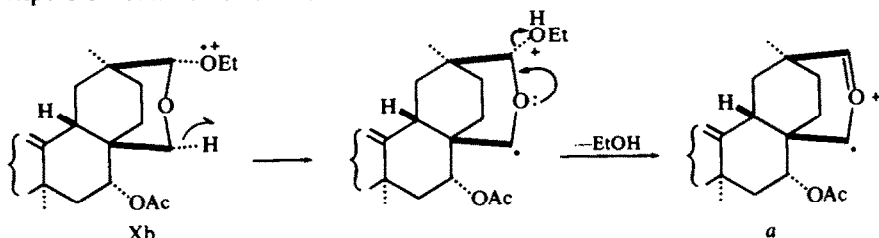


Fig. 2

these bonds are staggered and an axial 30α -EtO group is relatively unhindered (the nearest non-bonded neighbour is H-21 β at 2.2 Å). Since the ratio of cyclamigenins A¹:C is ca. 30:1, it is reasonable to assume that cyclamigenin C is the axial, 30α -epimer (IXb) and that cyclamigenin A¹ is the equatorial, 30β -isomer (IXa). There is some slight spectroscopic evidence in favour of these assignments. Thus, in the mass spectrum of cyclamigenin C diacetate M — EtOH (a) accounts for the base peak whereas this species is much less abundant (13 % relative intensity) in the case of cyclamigenin A¹ diacetate. It seems plausible that such a fragmentation would occur more easily for the axial, 30α -isomer where a preliminary hydrogen-transfer could lead to the expulsion of an ethanol molecule:



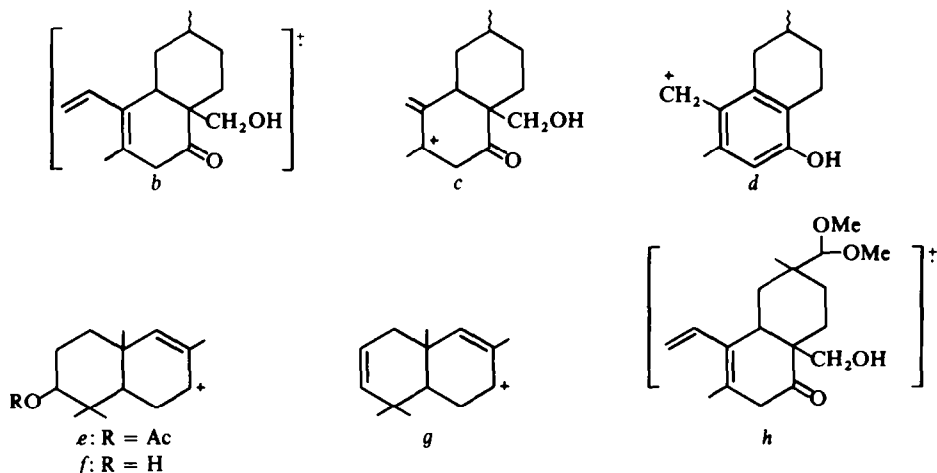
The hydrogen-transfer step is not possible in the case of the equatorial, 30β -epimer. Cyclamigenins A¹ and C are, therefore, 30β - and 30α -ethoxy-28,30-epoxyolean-12-en-3 β ,16 α -diol, respectively.

Both compounds are obviously artifacts although we have not been able to prepare them from cyclamiretin D (XI) which is unchanged when subjected to the conditions of the hydrolysis stage of the extraction,¹ viz treatment with aqueous ethanolic hydrochloric acid. With ethanolic hydrogen chloride cyclamiretin D yields the diethyl acetal (XIV).

Cyclamigenin A² is saturated to tetranitromethane and shows no selective absorption in the UV. Its IR spectrum, which is very similar to that of aegicerin (III),⁵ shows the presence of OH (3400 cm^{-1}), ketonic CO (1704 cm^{-1}), and ether oxygen (1028 and 890 cm^{-1}). It forms a monoacetate the NMR spectrum of which is similar to that of aegicerin acetate (IV) (e.g. the methylene group on the ether bridge appears as an AB quartet, $J = -8\text{ Hz}$, centred at $\tau\ 6.28$) except that it indicates the presence of only six skeletal Me groups one of which is secondary (doublet at $\tau\ 9.08$, $J = 7\text{ Hz}$). Cyclamigenin A², therefore, is 30(or 29)-noraegicerin (V). Confirmation of this structure is found in the mass spectrum of the acetate (VI) which is very similar to that¹ of aegicerin acetate (IV) except that the fragments containing ring E occur at 14 a.m.u. lower. Thus, the molecular ion appears at $m/e\ 484$, $\text{C}_{31}\text{H}_{48}\text{O}_4$, and the D/E-ring fragments *b*, *c*, and *d* occur at $m/e\ 234$, 221, and 189 (base peak), respectively.

Cyclamigenin D, which shows no selective absorption in the UV, absorbs in the IR at 3500 (O—H stretch) and 1707 cm^{-1} (ketonic C=O stretch). The IR spectra of both the alcohol and its acetate show a group of ether bands⁶ near 1100 cm^{-1} and one at 890 cm^{-1} (tertiary ether⁷) together with a peak at 2825 cm^{-1} indicative⁸ of the presence of MeO. The presence of the latter is confirmed by the NMR spectrum of cyclamigenin D acetate which shows a strong signal at $\tau\ 6.52$ the integrated intensity of which corresponds to two MeO groups. The spectrum shows only six angular Me groups. A singlet at $\tau\ 5.76$ (O—CH—O), an AB quartet ($J = -8\text{ Hz}$) centred at $\tau\ 6.3$ (CH_2O), and a doublet ($J = -16\text{ Hz}$) at $\tau\ 7.3$ (one proton of CH_2CO) suggested

that cyclamigenin D is the dimethyl acetal (VII) of cyclamigenin B (I). The mass spectra of cyclamigenin D (VII) and its acetate (VIII) are consistent with the proposed structure in that the molecular ions occur at m/e 516, $C_{32}H_{52}O_5$, and 558, $C_{34}H_{54}O_6$, respectively; A/B-ring fragments⁹ are observed at m/e 249 (*e*), 207 (*f*), and 189 (*g*); and D/E-ring fragments⁹ (e.g. the retro-diene species *h*) carry the acetal group. The skeletal fragments are all of low abundance and ca. 30% of the total ion current in both spectra is carried by the expected¹⁰ α -cleavage species, $MeOCH^+OMe$, of m/e 75.



Acid hydrolysis of cyclamigenin D yields a mixture of products (presumably liberation of the aldehyde is accompanied by some cleavage of the ether ring and elimination of C-28 as formaldehyde) but acetolysis followed by mild, chromic acid oxidation proceeds smoothly to give 3 β ,28-diacetoxy-16-ketoolean-12-en-30-oic acid (XV), identical with a sample prepared³ from cyclamiretin D diacetate (XIII). Finally, brief treatment of cyclamigenin B acetate (II)¹ with anhydrous methanol containing boron trifluoride etherate affords, in poor yield, the dimethyl acetal (VIII) which is identical with cyclamigenin D acetate.

Cyclamigenin D seems obviously an artifact although the only contact between cyclamigenin B and methanol was during crystallization of the acetate (II) in the presence of chloroform and no trace of cyclamigenin D acetate was detected (TLC) after prolonged treatment of the acetate (II) with refluxing chloroform-methanol.

Cyclamigenin E, only trace amounts of which were isolated,¹ was not obtained pure and was not investigated.

EXPERIMENTAL

Specific rotations were determined for 1% solns in $CHCl_3$ at 31° with a Perkin-Elmer 141 Polarimeter. M.p.s were determined in vacuum-sealed capillaries in an electrically heated aluminium block and are corrected. IR spectra were recorded for KBr discs on a Grubb-Parsons 'Spectromaster', NMR spectra in $CDCl_3$ on a Varian HR-60A instrument with TMS as internal standard, and mass spectra on A.E.I. MS-9 instruments operating at 70 eV with direct-insertion probes; ion source temp 190–230°. The constitutions of all important ions were confirmed by high-resolution measurements. Analytical samples were dried for 60 hr at $78^\circ/10^{-2}$ mm.

Cyclamigenin A¹ (IXa)

Cyclamigenin A¹ diacetate, isolated as described previously,¹ had m.p. 231–233°, $[\alpha]_D +15 \pm 1^\circ$, ν_{\max} 1735(s), 1235(s), 1000(s), 826(m), 814(w), 797(w) cm^{-1} , τ 9.12 (Me), 9.08 (4Me), 8.79 (tr, $J = 7$ Hz, Me), 8.77 (Me), 8.01 (Ac), 7.97 (Ac), 6.77 (d, $J = -12$ Hz, H-28), 6.42 (d, $J = -12$ Hz, H-28), 6.41 (m, H-16), 6.40 (q, OCH_2Me), 5.63 (s, H-30), 5.55 (m, H-3), 4.65 (m, H-12). (Found: C, 74.3; H, 9.7; EtO, 7.7. Calc. for $\text{C}_{36}\text{H}_{56}\text{O}_6$: C, 73.9; H, 9.65; EtO, 7.7%). Mass spectrum (m/e)/% relative abundance 584 (M^+)/1, 539/4, 538/13, 392/100, 377/50, 334/1, 321/1, 288/3, 274/3, 261/2, 249/2, 228/4, 189/4. Saponification of the acetate with 5% methanolic KOH furnished cyclamigenin A¹ as platelets, m.p. 216–217° (from MeOH), $[\alpha]_D +42^\circ$, ν_{\max} 3401(s), 1000(s), 831(m), 817(w), 802(w) cm^{-1} . (Found: C, 75.4; H, 10.5. Calc. for $\text{C}_{32}\text{H}_{52}\text{O}_4 \cdot \text{O} \cdot 5\text{MeOH}$: C, 75.5; H, 10.5%). Cyclamigenin A¹ is recovered quantitatively after 24 hr in refluxing 10% methanolic KOH. Mass spectrum (m/e)/% relative abundance 500 (M^+)/1, 482/2, 455/12, 454/31, 357/27, 292/3, 246/12, 207/17, 189/8, 112/100.

Cyclamigenin C (IXb)

Cyclamigenin C diacetate, isolated as described previously,¹ had m.p. 222–223°, $[\alpha]_D -125 \pm 2^\circ$, ν_{\max} 1735(s), 1235(s), 1000(s), 827(m), 814(w), 798(w) cm^{-1} , τ 9.14 (3Me), 9.08 (2Me), 8.81 (tr, $J = 7$ Hz, Me), 8.74 (Me), 8.01 (Ac), 7.95 (Ac), 6.79 (d, $J = -12$ Hz, H-28), 6.28 (d, $J = -12$ Hz, H-28), 6.50 (q, OCH_2Me), 6.4 (m, H-16), 5.63 (s, H-30), 5.55 (m, H-3), 4.65 (m, H-12). (Found: C, 74.1; H, 9.8; EtO, 8.2. Calc. for $\text{C}_{36}\text{H}_{56}\text{O}_6$: C, 73.9; H, 9.65; EtO, 7.7%). Mass spectrum (m/e)/% relative abundance 584 (M^+)/2, 539/40, 538/100, 321/1, 288/7, 274/7, 261/1, 249/2, 228/6, 189/5. Saponification of the acetate with 5% methanolic KOH gave cyclamigenin C as platelets, m.p. 218–219° (from MeOH aq), $[\alpha]_D -114^\circ$, ν_{\max} 3401(s), 998(s), 830(m), 815(w), 800(w) cm^{-1} . (Found: C, 76.5; H, 10.3. Calc. for $\text{C}_{32}\text{H}_{52}\text{O}_4 \cdot \text{C}, 76.75$; H, 10.45%). Cyclamigenin C is recovered quantitatively after 24 hr in refluxing 10% methanolic KOH. Mass spectrum (m/e)/% relative abundance 500 (M^+)/2, 482/3, 455/14, 454/27, 357/79, 292/4, 246/8, 207/16, 189/7, 112/100.

Cyclamiretin D monoacetate (XII)

(a) A soln of cyclamigenin A¹ diacetate (27 mg) in THF (10 ml) containing 10% HCl aq (3 ml) was left overnight at room temp and then evaporated to dryness at 30°. Crystallization of the residue from CHCl_3 —MeOH yielded cyclamiretin D monoacetate (12 mg), m.p. 237–239°, $[\alpha]_D +36^\circ$, ν_{\max} 3580(s), 2702(w), 1730(s, br), 1245(s) cm^{-1} , identical (mixed m.p. and IR spectrum) with the samples prepared from cyclamigenin C and cyclamiretin D acetates. (Found: C, 72.1; H, 10.1. Calc. for $\text{C}_{32}\text{H}_{50}\text{O}_5 \cdot \text{MeOH}$: C, 72.5; H, 10.0%).

(b) Similar treatment of cyclamigenin C diacetate (25 mg) gave cyclamiretin D monoacetate (9 mg), m.p. 237–239°, $[\alpha]_D +34^\circ$, identical (mixed m.p. and IR spectrum) with samples prepared from cyclamigenin A¹ and cyclamiretin D diacetates.

(c) Similar treatment of cyclamiretin D diacetate^{3,4} (50 mg) afforded cyclamiretin D monoacetate (18 mg), m.p. 237–239°, $[\alpha]_D +36^\circ$, identical (mixed m.p. and IR spectrum) with samples prepared from cyclamigenin A¹ and C diacetates.

Cyclamiretin D diethyl acetal (XIV)

Cyclamiretin D (50 mg) in dry EtOH (25 ml) saturated with HCl was stored (2 days) at 0°, the soln evaporated to dryness at 0–5°, and the residue crystallized from MeOH to yield the diethyl acetal (15 mg), m.p. 340–343°, $[\alpha]_D +119^\circ$, ν_{\max} 3410(s), 1105(s), 1087(s), 827(m) cm^{-1} . (Found: C, 72.8; H, 10.8. Calc. for $\text{C}_{34}\text{H}_{58}\text{O}_5 \cdot \text{MeOH}$: C, 72.6; H, 10.8%). Acetylation of the alcohol, with Ac_2O in pyridine at room temp, furnished the corresponding diacetate, m.p. 318–320° (from hexane—EtOH), $[\alpha]_D +90^\circ$. (Found: C, 72.1; H, 9.8. Calc. for $\text{C}_{38}\text{H}_{62}\text{O}_7$: C, 72.3; H, 9.9%).

Cyclamiretin D (100 mg) in EtOH (50 ml) was refluxed (3 hr) with 10% HCl aq (50 ml). The product (97 mg), m.p. 239–241°, $[\alpha]_D +35^\circ$, precipitated by addition of water (100 ml), was identical (mixed m.p. and IR spectrum) with the starting material and thin-layer chromatography showed no trace of cyclamigenins A¹ or C.

Cyclamigenin A² (V)

The crude cyclamigenin B acetate (790 mg) from fraction C of the mixture of sapogenins¹ was dissolved in benzene and chromatographed on alumina (40 g). Elution with benzene (1.4 l) gave a fraction (390 mg) crystallization of which from CHCl_3 —MeOH furnished cyclamigenin A² acetate as fine needles, m.p. 231–233°, $[\alpha]_D -2^\circ$, ν_{\max} 1735(s), 1705(s), 1245(s), 1023(s), 896(s) cm^{-1} , τ 9.14 (2Me), 9.09 (Me), 9.08 (d,

$J = 7$ Hz, Me), 8.76 (Me), 8.37 (Me), 7.96 (Ac), 7.25 (d, $J = -15$ Hz; H-15), 6.48 (d, $J = -8$ Hz, H-28), 6.08 (d, $J = -8$ Hz, H-28), 5.5 (m, H-3). (Found: C, 74.6; H, 10.2. Calc. for $C_{31}H_{48}O_4$: C, 74.3; H, 10.1%). Mass spectrum (m/e)/% relative abundance 484 (M^+)/12, 483/9, 482/24, 315/37, 249/8, 234/27, 221/41, 219/42, 203/64, 191/49, 190/58, 189/100. Saponification of the acetate yielded cyclamigenin A² as feathery needles, m.p. 208–209° (from MeOH), $[\alpha]_D -17^\circ$, ν_{max} 3400(s), 1704(s), 1028(s), 890(s) cm^{-1} . (Found: C, 76.2; H, 10.5. Calc. for $C_{29}H_{46}O_3$: C, 75.9; H, 10.6%).

Further elution of the column with benzene (1.5 l.) gave a mixture (380 mg) of cyclamigenin A² and B acetates.

Cyclamigenin D (VII)

(a) Cyclamigenin D acetate, isolated as described previously,¹ had m.p. 272–274°, $[\alpha]_D +3 \pm 1^\circ$, ν_{max} 2825(m), 1735(s), 1707(s), 1143(s), 1105(s), 1071(s), 1002(s), 890(m) cm^{-1} , τ 9.14 (4Me), 8.98 (Me), 8.77 (Me), 7.99 (Ac), 7.3 (d, $J = -16$ Hz, H-15), 6.52 (2MeO), 6.56 (d, $J = -8$ Hz, H-28), 6.15 (d, $J = -8$ Hz, H-28), 5.76 (s, H-30), 5.5 (m, H-3). (Found: C, 73.5; H, 9.7. Calc. for $C_{34}H_{54}O_6$: C, 73.1; H, 9.7%). Mass spectrum (m/e)/% relative abundance 558 (M^+)/0.2, 526/0.5, 308/0.8, 249/0.3, 189/1, 75/100. Saponification of the acetate gave cyclamigenin D as needles, m.p. 325–327°, $[\alpha]_D -5^\circ$, ν_{max} 2825(m), 1707(s), 1143(s), 1105(s), 1003(s), 889(s) cm^{-1} . (Found: C, 74.4; H, 10.1. Calc. for $C_{32}H_{52}O_5$: C, 74.4; H, 10.1%). Mass spectrum (m/e)/% relative abundance 516 (M^+)/0.2, 526/0.5, 308/0.8, 249/0.3, 189/1, 75/100.

(b) A suspension of cyclamigenin B acetate (200 mg) in anhydrous MeOH (3 ml) was stirred (1 hr) at room temp with BF_3 -etherate (0.5 ml), the mixture diluted with MeOH (5 ml), filtered, and the solid washed with cold MeOH (5 ml). Preparative TLC (alumina/benzene) afforded cyclamigenin B acetate (75 mg), m.p. 306–308°, $[\alpha]_D -9^\circ$, and VIII (12 mg), m.p. 271–273°, $[\alpha]_D +4^\circ$, identical (mixed m.p. and IR spectrum) with cyclamigenin D acetate. Alkaline hydrolysis of the acetate furnished the corresponding alcohol (VII), m.p. 324–326°, $[\alpha]_D -5^\circ$, identical (mixed m.p. and IR spectrum) with cyclamigenin D.

Acetolysis of cyclamigenin D

Cyclamigenin D (50 mg) in Ac_2O (3 ml) was heated (30 min) at 115–120° with *p*-toluenesulphonic acid (30 mg), the mixture poured into ice-water (25 ml), and worked up through ether. A soln of the product in AcOH (2 ml) was left (30 min) at room temp with CrO_3 (25 mg) in AcOH (2 ml), the mixture was diluted with water (10 ml), the precipitated solid washed with water, and crystallized from MeOH/aq to yield needles (21 mg) of 3 β ,28-diacetoxy-16-keto-olean-12-en-30-oic acid, m.p. 275–277° (dec), $[\alpha]_D +11^\circ$, identical (mixed m.p. and IR spectrum) with a sample prepared³ from cyclamiretin D diacetate.

Acknowledgements—We are indebted to the Shiftung Volkswagenwerk for the purchase of the mass spectrometer, to Mr. M. Hoog (C. G. Van Tubergen Ltd., Haarlem) for generous gifts of authenticated plant material, and to Prof. C. Djerassi and Dr. J. K. MacLeod (Stanford) for some of the low-resolution spectra.

REFERENCES

- 1 Part III, R. Ó Dorchai and J. B. Thomson, *Tetrahedron* **24**, 1377 (1968).
- 2 A. R. H. Cole and D. W. Thornton, *J. Chem. Soc.* 1332 (1957).
- 3 R. Tschesche, H. Striegler and H. W. Fehlhaber, *Liebigs Ann.* **691**, 165 (1966).
- 4 D. H. R. Barton, A. Hameed and F. J. McGhie, *J. Chem. Soc.* 5176 (1962); R. Tschesche, F. Inchaurredo, and G. Wulff, *Liebigs Ann.* **680**, 107 (1964).
- 5 K. Venkateswara Rao, *Tetrahedron* **20**, 973 (1964).
- 6 E. D. Bergmann and S. Pinchas, *Rec. Trav. Chim.* **71**, 161 (1952); H. Tschamler and R. Leutner, *Monatsh* **83**, 1502 (1952).
- 7 A. D. Cross, *Introduction to Practical Infra-red Spectroscopy* p. 60. Butterworths, London (1960).
- 8 H. B. Henbest, G. D. Meakins, B. Nicholls and A. A. Wagland, *J. Chem. Soc.* 1462 (1957).
- 9 H. Budzikiewicz, J. M. Wilson and C. Djerassi, *J. Am. Chem. Soc.* **85**, 3688 (1963); J. Karliner and C. Djerassi, *J. Org. Chem.* **31**, 1945 (1966).
- 10 H. Budzikiewicz, C. Djerassi and D. H. Williams, *Mass Spectrometry of Organic Compounds* p. 259. Holden-Day, San Francisco (1967).